

RNA Sequencing

Sample Submission Guidelines - RNA Sequencing

Step 1: Prepare your samples according to the table below. Please contact us at support@cofactorgenomics.com for Custom Project requirements.

- RNA is DNase-treated and is resuspended in nuclease-free water.
- If possible, assess RNA quality on a fragment analyzer (e.g. Agilent bioanalyzer).
- Measure RNA concentration using a fluorescent based method (e.g. Qubit). If using a NanoDrop to assess sample
 purity, the A260/280 ratio should be >1.9 and A260/230 ratio >2.0

Product Name	Library Type	Minimum Amount of Tissue	Recommended RNA Quantity‡	Preferred Concentration (Minimum Concentration)
mRNAble	Standard mRNA Sequencing (poly-A enrichment)	10-30 mg tissue or 10 ⁶ mammalian cell pellet	≥500 ng	50 ng/μl (5 ng/μl)
Total RNAble*	Total RNA Sequencing (whole transcriptome/ rRNA depleted)	10-30 mg tissue or 10 ⁶ mammalian cell pellet	≥500 ng	100 ng/μl (15 ng/μl)
picoRNA	Low-input RNA Sequencing (based on Poly-A enrichment)	Contact us	≥100 pg or equivalent cell number (shipped in lysis buffer)	1 ng/µl (100 pg/µl)
FFPExact*	Fragmented RNA Sequencing (whole transcriptome/ rRNA depleted)	2 x 10 μm FFPE sections ‡	≥500 ng	100 ng/µl (30 ng/µl)
miRNAble	miRNA enrichment	10-30 mg tissue, 5 mL bacterial culture pellet, or 10 ⁶ mammalian cell pellet	≥l µg	100 ng/μl (20 ng/μl)
RNAccess*	Low-input/Low-quality (targeted) RNA Sequencing	2 x 10 μm FFPE sections [‡] or 5-10 mg tissue	≥40 ng	20 ng/µl (5 ng/µl)

‡Some tissues such as epidermal tissue may require larger amounts.

*Total RNAble and FFPExact are specifically for RNA derived from human, mouse, and rat samples; RNAccess is only for human samples. All other products/catalog numbers are amenable to any organism.

Step 2: Label your samples.

- RNA is submitted in 1.5 ml micro-centrifuge tubes clearly labeled with sample names matching the Sample Requisition Form (below), preferably tubes are sealed with parafilm.
- Tissue, cell pellets, and FFPE sections are submitted in 1.5 ml micro-centrifuge tubes clearly labeled with sample names matching the Sample Submission Form (linked below), preferably tubes are sealed with parafilm. We suggest removing as much excess block material from FFPE samples as possible.

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Sample Submission Guidelines - RNA Sequencing, continued

Step 3: Package your samples.

- Fresh tissues and extracted RNA are frozen and will be shipped on plenty of dry ice. We recommend these samples be snap-frozen on dry ice/ethanol or in liquid nitrogen as soon as possible after harvest or excision. Please avoid shipping samples on Fridays to keep samples from prolonged exposure to ambient temperature.
- Fresh tissue may also be preserved in RNAlater stabilization solution according to the manufacturers instructions.
- Enclose all samples within an interior box to keep them from physical contact with dry ice.

Step 4: Download the RNA Sequencing Sample Requisition Form from: <u>https://cofactorgenomics.com/sample-submission/</u>

• Complete this spreadsheet; be sure to include sample groupings required for group-wise analysis.

Step 5: Ship your materials.

Ship packages to: Cofactor Genomics Attn: Sample Submission 4044 Clayton Ave. St. Louis, MO 63110

Step 6: Complete the web form at <u>https://cofactorgenomics.com/sample-submission/</u> and upload your Sample Requisition Form at the address above. Please include your shipment's Tracking Information to ensure package monitoring.

International customers please refer to the "Guidelines for International Shipments" document on the sample submission page for additional shipping instructions.